CLEANING UP DIRTY DNA PREPS

(Modified from “current protocols” 2.3.3 by H. Judelson)

1. To DNA add 1 volume of: 2% CTAB, 100 mM Tris 8.0, 20 mM EDTA, 1.4 M NaCl (preferably at 55-65^{0}C). Mix by inversion. Incubate at 65^{0}C for 10 mm.

2. Add equal volume of 24:1 chloroform: isoamyl alcohol [or octanol]. Invert; vortex briefly (up to 30 sec).

3. Spin 5 min in mícrofuge to separate phases.

4. Save supernatant, being careful not to remove the interface (it desired to increase the yield, back extract the chloroform-interface with 150 µl of the 2% CTAB solution).

5. Add 1.1 volume of (preferably at 55-65^{0}C; this increases the speed of precipitation): 1% CTAB, 50 mM Tris 8.0,10 mM EDTA. Mix well by inversion. A precipitate should be visible.

6. Spin 5 min, 3k x g. If pellets are seen, discard the supematant.

7. Add to me pellet 0.5 ml of: 10 mM Tris 8.0, 0.1 mM EDTA, 1 M NaCl. Heat 30 mm, 65^{0}C to dissolve the pellet. Flick the tube to make sure the pellet is dissolved.

8. Add 0.6 vol. isopropanol (0.3 ml). Invert several times. Spin 8 mm in microfuge. Discard supernatant.

9. AdcIO.5 ml 70% ethanol, spin 1 minute in microfuge, and discard pellet.

10. Add TE and heat 65^{0}C for 30 mm.