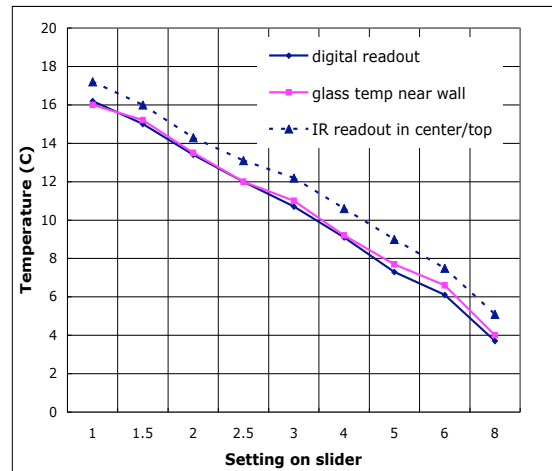


## Use of the microcooler for ligations

1. This was purchased for ligations, although other applications are possible.
2. It takes <10 minutes to cool down; turn on just before use, turn off when done.
3. Put your tubes in the black rack (don't fill the chamber with water).
4. The temperature in the top/middle of the rack is slightly higher than the digital readout, which measures the temperature at the chamber wall (see below). The temperature in the middle of the chamber is probably in between. So for 14°C or 16°C ligations, aim for about 13 or 15°C on the readout (slider positions 1.5 and 2, respectively). The exact temperature is not critical.

Recorded temperatures (°C) ( may vary slightly depending on room temperature?)			
Setting (slider)	Instrument readout	Thermometer (top at wall)*	IR gauge (rack center/top)*
1	16.2	16.0	17.2
1.5	15.0	15.2	16.0
1.75	14.4	14.8	14.9
2	13.4	13.5	14.3
2.5	12.0	12.0	13.1
3	10.7	11.0	12.2
4	9.1	9.2	10.6
5	7.3	7.7	9.0
6	6.1	6.6	7.5
8	3.7	4.0	5.1

\*readings may not be 100% accurate.



Supplementary information: Most experiments use T4 DNA Ligase, which is most active at 25°C. However, to perform efficient ligations with cohesive-ended fragments ("sticky ends"), the optimal enzyme temperature is balanced with the melting temperature of the ends. All ligations work fine at low temperatures (4°C), but go faster at higher temperatures, as long as the  $T_m$  is not exceeded.

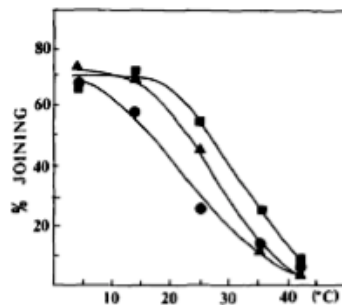


Figure 2. Temperature dependence of the joining of DNA termini produced by *EcoRI* (▲), *PstI* (■) and *HaeIII* (●).

*EcoRI* sticky ends are 100% AT; *PstI* sticky ends are 50% AT; *HaeIII* ends are blunt. Hence *EcoRI* ends ligate most efficiently at the lowest temperature, *HaeIII* at the highest, *PstI* is intermediate. From Ferretti, 1981.